

# ZERNIKE INSTITUTE COLLOQUIUM

Thursday, June 21<sup>st</sup>, 2012

16:00h, Lecture Hall: 5111.0080

Coffee and cakes from 15:30h

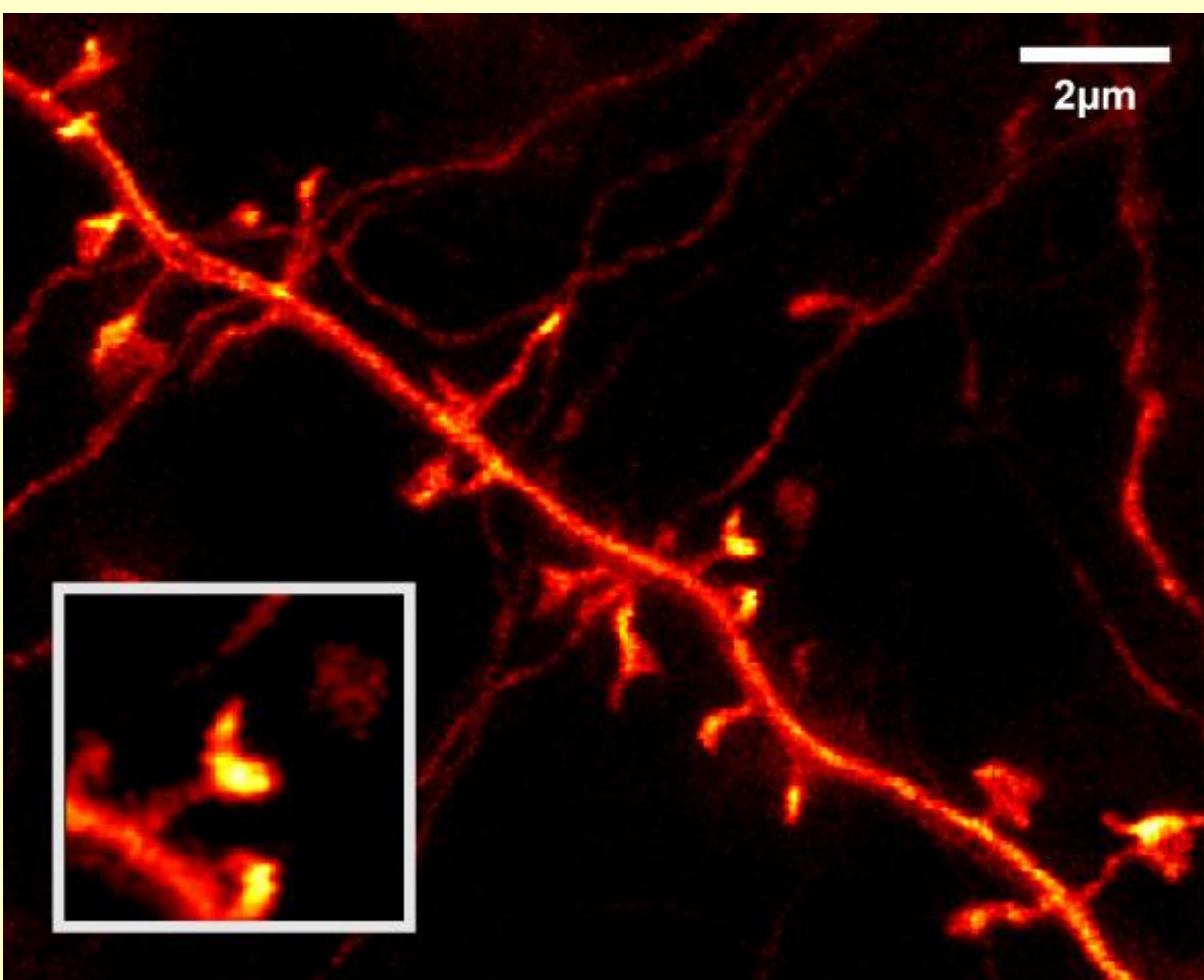
## NANOSCOPY WITH FOCUSED LIGHT

**Stefan W. Hell**  
**Max Planck Institute for  
Biophysical Chemistry**  
**Göttingen, Germany**



In STED microscopy<sup>1</sup>, fluorescent features are switched off by the STED beam, which confines the fluorophores to the ground state everywhere in the focal region except at a subdiffraction area of extent  $d \approx \lambda / (2NA\sqrt{1 + I/I_s})$ .

In RESOLFT microscopy<sup>2,3</sup>, the principles of STED have been expanded to fluorescence on-off-switching at low intensities  $I$ , by resorting to molecular switching mechanisms that entail low switching thresholds  $I_s$ . An  $I_s$  lower by many orders of magnitude is provided by reversibly switching the fluorophore to a long-lived dark (triplet) state<sup>2-4</sup> or between a long-lived 'fluorescence activated' and 'deactivated' state<sup>2,5</sup>.



These alternative switching mechanisms entail an  $I_s$  that is several orders of magnitude lower than in STED. In imaging applications, STED/RESOLFT enables fast recordings and the application to living cells, tissues, and even living animals<sup>6,7</sup>.

Starting from the basic principles of nanoscopy we will discuss recent developments<sup>8,9</sup> with particular attention to RESOLFT and the recent nanoscale imaging of the brain of living mice<sup>7</sup> by STED.

*STED movie from a living mouse brain Neuron recorded from the molecular layer of the somatosensory cortex of a living transgenic mouse expressing YFP with resolution < 68 nm, showing moving dendritic spines.*

1. Hell, S. W. & Wichmann, J. Breaking the diffraction resolution limit by stimulated-emission - stimulated-emission-depletion fluorescence microscopy. *Opt Lett* **19**, 780-782, doi:10.1364/OL.19.000780 (1994).
2. Hell, S. W. Toward fluorescence nanoscopy. *Nat Biotechnol* **21**, 1347-1355 (2003).
3. Hell, S. W., Jakobs, S. & Kastrup, L. Imaging and writing at the nanoscale with focused visible light through saturable optical transitions. *Appl Phys A* **77**, 859-860 (2003).
4. Hell, S. W. Far-Field Optical Nanoscopy. *Science* **316**, 1153-1158 (2007).
5. Hofmann, M., Eggeling, C., Jakobs, S. & Hell, S. W. Breaking the diffraction barrier in fluorescence microscopy at low light intensities by using reversibly photoswitchable proteins. *PNAS* **102**, 17565-17569 (2005).
6. Rankin, B. R. et al. Nanoscopy in a Living Multicellular Organism Expressing GFP. *Biophys J* **100**, L63 - L65 (2011).
7. Berning, S., Willig, K. I., Steffens, H., Dibaj, P. & Hell, S. W. Nanoscopy in a Living Mouse Brain. *Science* **335**, 551 (2012).
8. Grotjohann, T. et al. Diffraction-unlimited all-optical imaging and writing with a photochromic GFP. *Nature* **478**, 204-208 (2011).
9. Brakemann, T. et al. A reversibly photoswitchable GFP-like protein with fluorescence excitation decoupled from switching. *Nat Biotechnol* **29**, 942-947 (2011).



university of  
 groningen

zernike institute for  
 advanced materials